



Clinical results CAR-T for mantle cell lymphoma: what's next?

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Disclosures

Research: Pharmacyclics, Acerta Pharma, Kite Pharma, Velosbio, Inc, InnoCare, BeiGene, Loxo Oncology, AstraZeneca, Genetech, Genmab, BeiGene Aus Pty Ltd, Incyte, Lilly, BeiGene LTD, Juno Therapeutics, Janssen, Vincerx, Pharmacyclics, Nurix Therapeutics, AbbVie, Inctye, Bantam Pharmaceutical, LLC, Oncternal Therapeutics

Honoraria: AstraZeneca, BeOne, CAHON,, East Virginia Medical School,, Janssen, Kite Pharma, Mayo Clinic, MJH Life Sciences, Merck,, Pfizer,, PromCon S.R.E., Research to Practice, Studio ER Congressi, Medscape/WebMD, VJHemonc

Consultancy: AstraZeneca, Galapagos NV, Genmab, InnoCare, Janssen, Kite Pharma, Lilly, Merck, PER, Pepromene Bio, Oncternal

Five-Year Outcomes of Patients With Relapsed or Refractory Mantle Cell Lymphoma Treated With Brexucabtagene Autoleucel in ZUMA-2 Cohorts 1 and 2

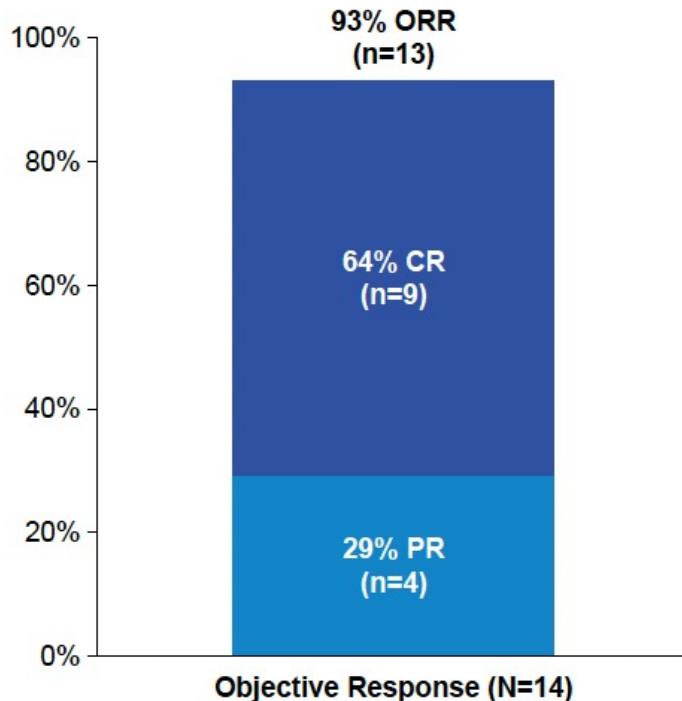
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¹⁸University of Rochester School of Medicine, Rochester, NY, USA; and ¹⁹University of Rochester School of Medicine, Rochester, NY, USA

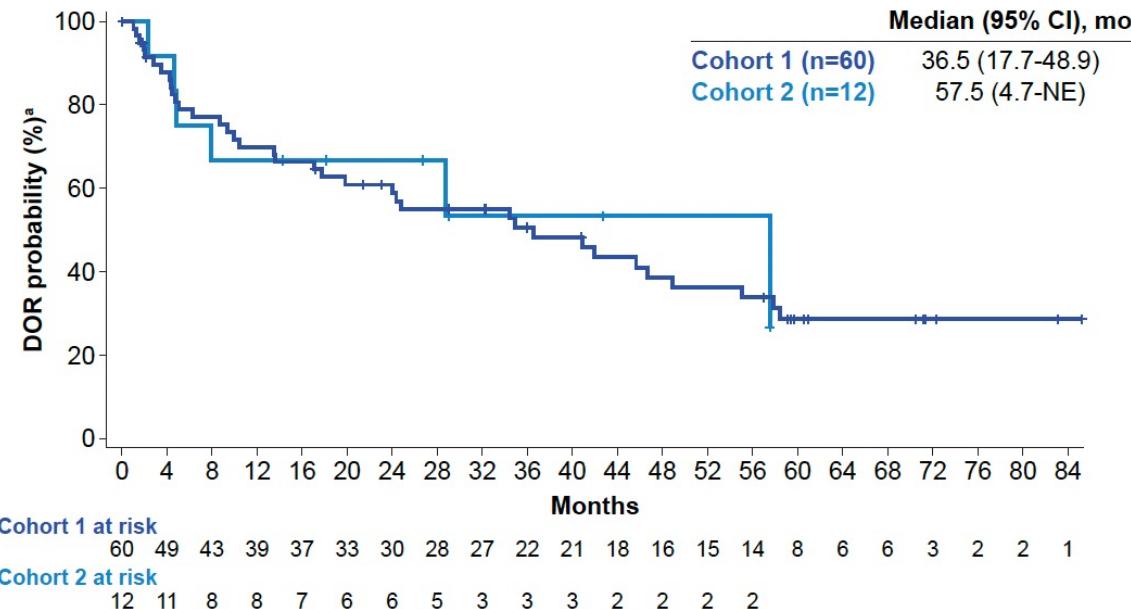
Best Objective Response by IRRC for Cohort 2 Primary Analysis



- In Cohort 2 primary analysis, ORR was 93% (95% CI, 66.1-99.8); 64% of patients had a CR and 29% had a PR
 - No patients had stable disease or progressive disease
 - One patient was not assessed at the time of analysis

CR, complete response; IRRC, independent radiology review committee; ORR, objective response rate; PR, partial response.

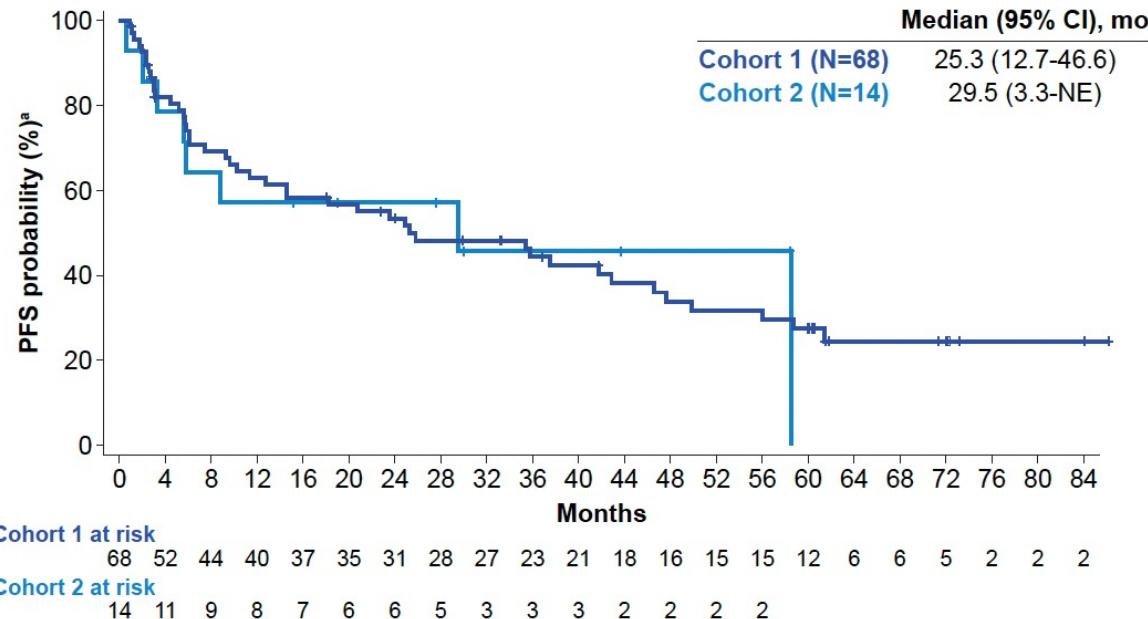
Duration of Response in ZUMA-2 5-Year Outcomes



- In Cohort 1, median investigator-assessed DOR was 36.5 months (95% CI, 17.7-48.9; n=60) with 17 patients in ongoing response at data cutoff, all CR
- In Cohort 2, median DOR was 57.5 months (95% CI, 4.7-NE; n=12) with 3 patients in ongoing response at data cutoff, all CR

^a Per investigator assessment. CR, complete response; DOR, duration of response; NE, not estimable.

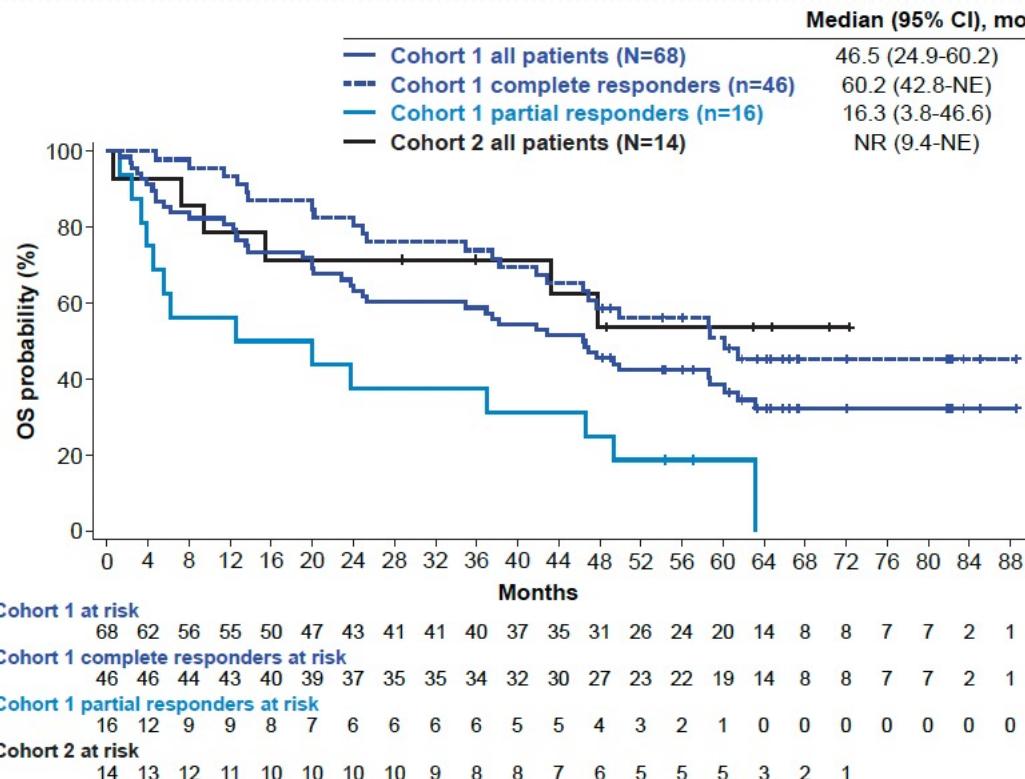
Progression-Free Survival in ZUMA-2 5-Year Outcomes



- Median investigator-assessed PFS was 25.3 months (95% CI, 12.7-46.6; N=68) and 54-month PFS rate was 32% (95% CI, 20.0-44.2) in Cohort 1
- In Cohort 2, median PFS was 29.5 months (95% CI, 3.3-NE) and 54-month PFS rate was 46% (17.3-70.5; N=14)

^a Per investigator assessment. NE, not estimable; PFS, progression-free survival.

Overall Survival in ZUMA-2 5-Year Outcomes



- In Cohort 1, the median OS was 46.5 months (95% CI, 24.9-60.2) and 60-month OS rate was 39% (95% CI, 26.7-50.1)
- In Cohort 2, median OS was not reached (95% CI, 9.4-NE) and 60-month OS rate was 54% (95% CI, 23.8-76.2)

NE, not estimable; NR, not reached; OS, overall survival.

Lisocabtagene maraleucel in R/R MCL: primary analysis results from the MCL cohort of the single-arm, multicenter, seamless design TRANSCEND NHL 001 study

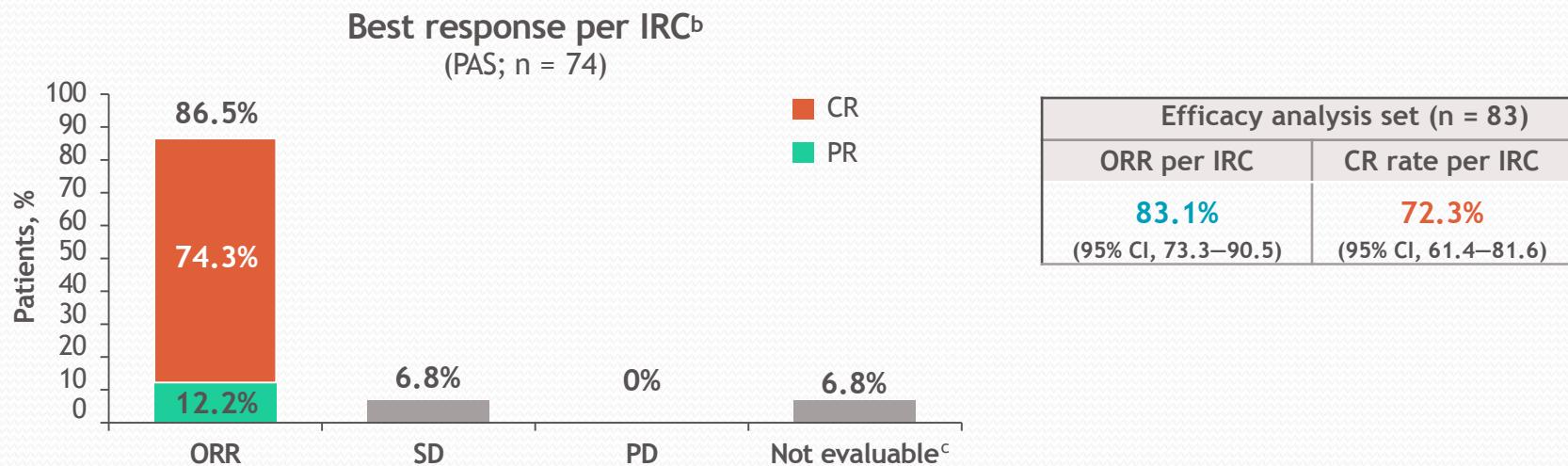
Michael Wang,¹ Tanya Siddiqi,² Leo I. Gordon,³ Manali Kamdar,⁴ Matthew Lunning,⁵ Alexandre V. Hirayama,⁶ Jeremy S. Abramson,⁷ Jon Arnason,⁸ Nilanjan Ghosh,⁹ Amitkumar Mehta,¹⁰ Charalambos Andreadis,¹¹ Scott R. Solomon,¹² Ana Kostic,¹³ Christine Dehner,¹³ Ricardo Espinola,¹⁴ Lily Peng,¹³ Ken Ogasawara,¹⁵ Amy Chattin,¹³ M. Lia Palomba¹⁶

¹The University of Texas MD Anderson Cancer Center, Houston, TX, USA; ²City of Hope National Medical Center, Duarte, CA, USA; ³Northwestern University, Feinberg School of Medicine, Robert H. Lurie Comprehensive Cancer Center, Chicago, IL, USA; ⁴University of Colorado Cancer Center, Aurora, CO, USA; ⁵University of Nebraska Medical Center, Omaha, NE, USA; ⁶Fred Hutchinson Cancer Center, Seattle, WA, USA; ⁷Massachusetts General Hospital Cancer Center, Harvard Medical School, Boston, MA, USA; ⁸Beth Israel Deaconess Medical Center, Boston, MA, USA; ⁹Levine Cancer Institute, Atrium Health, Charlotte, NC, USA; ¹⁰University of Alabama at Birmingham, Birmingham, AL, USA; ¹¹University of California, San Francisco, San Francisco, CA, USA; ¹²Northside Hospital Cancer Institute, Atlanta, GA, USA; ¹³

¹⁶Memorial Sloan Kettering Cancer Center, New York, NY, USA

Liso-cel delivers high ORR and CR rates in patients with R/R MCL

- Primary (ORR) and key secondary (CR rate) per IRC efficacy endpoints^a were met based on the PAS (n = 74)
 - ORR: **86.5%** (95% CI, 76.5–93.3); CR rate: **74.3%** (95% CI, 62.8–83.8); $P < 0.0001$ for both
- Consistently high ORR and CR rate were observed in the efficacy analysis set (n = 83)
 - Median (range) time to first CR or PR was 0.95 (0.7–3.0) months

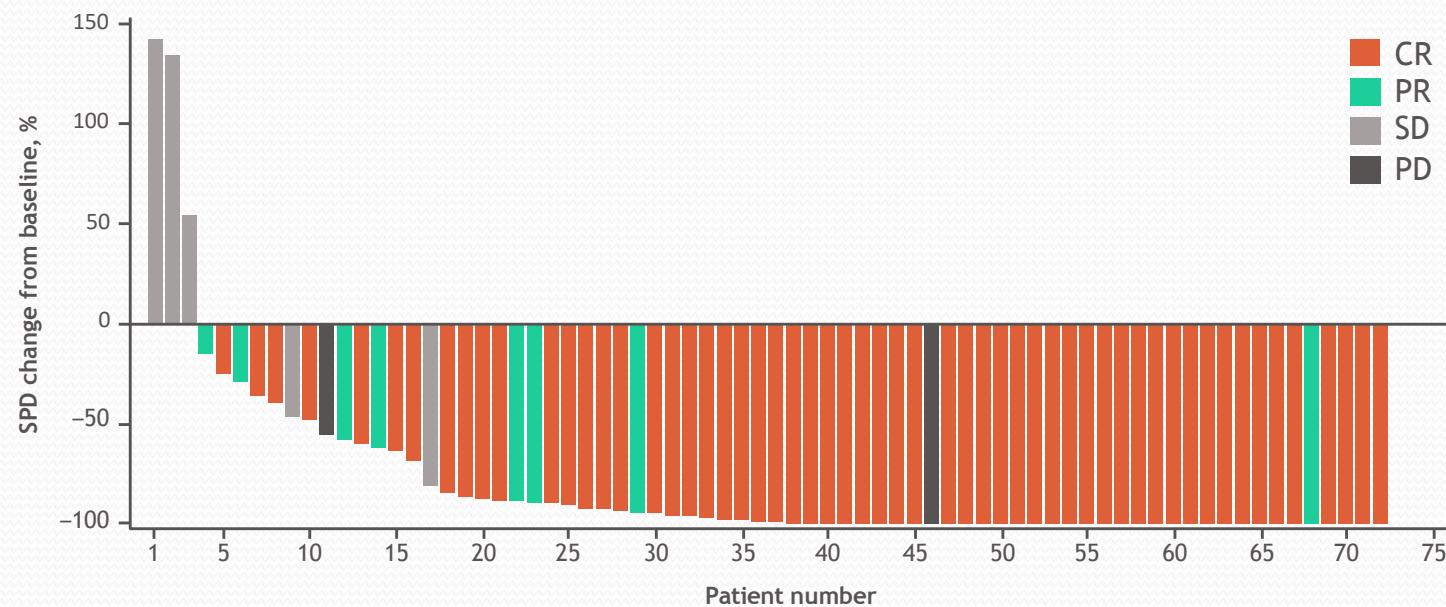


^aPrimary and key secondary efficacy hypotheses were tested hierarchically in the PAS in the order of the following: H_0 : ORR $\leq 40\%$, H_0 : CR rate $\leq 18\%$, one-sided p-value; ^bBest disease response per IRC by Lugano 2014 criteria from the time of liso-cel infusion until disease progression, end of study, start of another anticancer therapy, or HSCT; ^cNo postbaseline scans were submitted to IRC for evaluation. H_0 , null hypothesis.

Change in tumor burden and response status per IRC^a (efficacy analysis set)

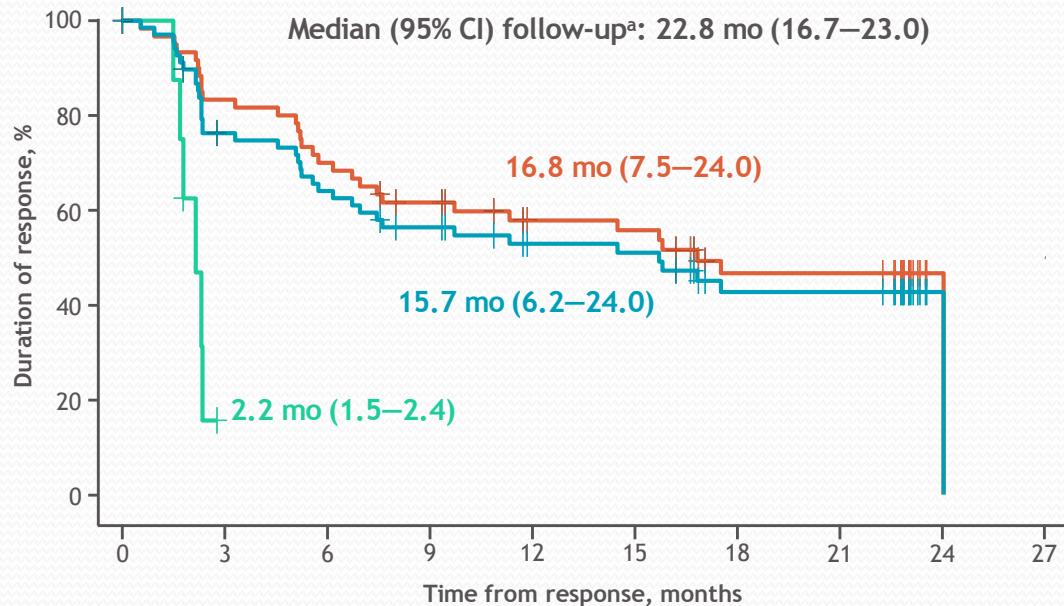
TRANSCEND NHL 001: MCL cohort

Most patients experienced a reduction in lymphadenopathy^a



^aMaximum change from baseline in SPD per IRC assessment for all patients in the efficacy analysis set with baseline and ≥ 1 postbaseline target lesion measurement. 11 patients were excluded due to lack of measurable target lesions at baseline (n = 5), or missing scans postbaseline (n = 6).

DOOR per IRC (efficacy analysis set)

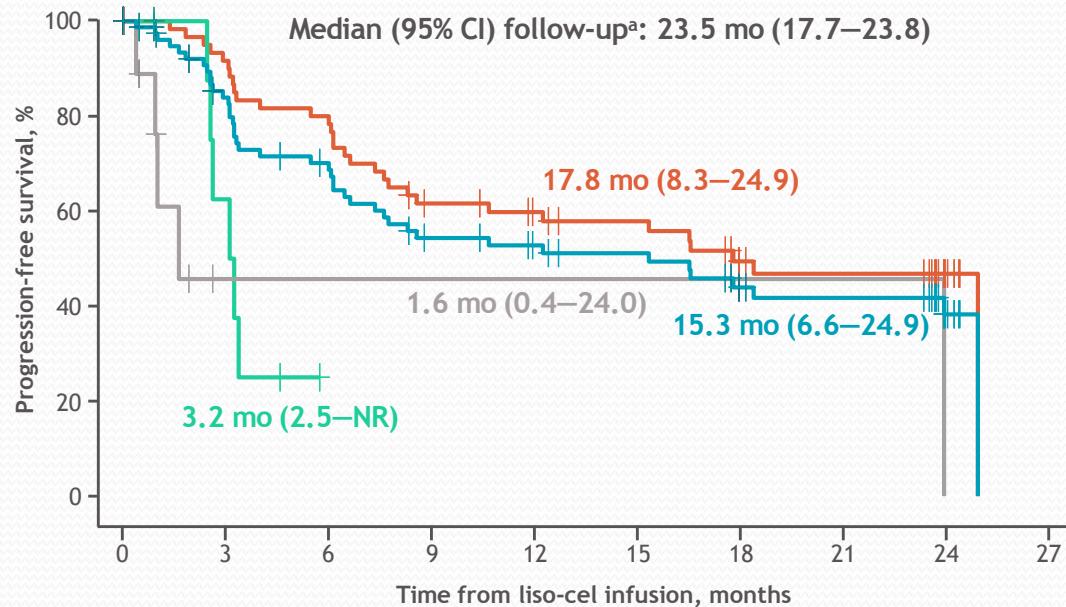


| | CR | PR | CR/PR |
|----|----|----|-------|
| 60 | 50 | 42 | 69 |
| 9 | 0 | 35 | 50 |
| 69 | 50 | 42 | 35 |

| | Continued response rate | |
|--|-----------------------------|------------------------------|
| | Responders (n = 69) | Patients with CR (n = 60) |
| 12-mo rate (95% CI) ^b | 52.9% (40.1–64.2) | 57.8% (44.2–69.2) |
| 18-mo rate (95% CI) ^b | 42.7% (29.9–54.9) | 46.7% (32.8–59.4) |

^aReverse Kaplan-Meier method was used to obtain median follow-up and its 95% CI; ^bKaplan-Meier method was used to obtain 2-sided 95% CI intervals.

PFS per IRC (efficacy analysis set)



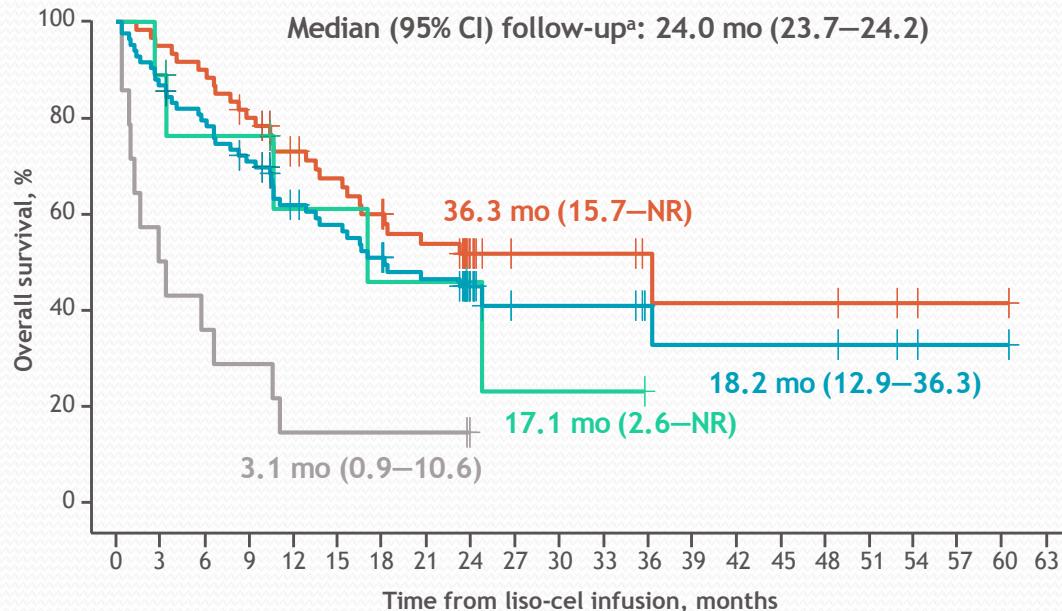
| | PFS rate | |
|----------------------------------|-----------------------------|-----------------------------|
| | Total (n = 83) | Patients with CR (n = 60) |
| 12-mo rate (95% CI) ^b | 52.8% (40.6–63.6) | 59.8% (46.3–71.0) |
| 18-mo rate (95% CI) ^b | 43.9% (31.8–55.4) | 49.4% (35.7–61.8) |

| | | | | | | | | | | |
|--------------|----|----|----|----|----|----|----|----|---|---|
| CR | 60 | 55 | 48 | 35 | 31 | 28 | 20 | 18 | 6 | 0 |
| PR | 9 | 5 | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 0 |
| Nonresponder | 14 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 |
| Total | 83 | 61 | 49 | 36 | 32 | 29 | 21 | 19 | 6 | 0 |

^aReverse Kaplan-Meier method was used to obtain median follow-up and its 95% CI; ^bKaplan-Meier method was used to obtain 2-sided 95% CI intervals.
NR, not reached.

Wang M, et al. ICML 2023 [Abstract #LBA3]

OS (efficacy analysis set)



| | OS rate | |
|--|-----------------------------|---------------------------------|
| | T otal (n = 83) | Patients with CR (n = 60) |
| 12-mo rate (95% CI) ^b | 61.8% (50.2–71.4) | 72.9% (59.6–82.5) |
| 18-mo rate (95% CI) ^b | 50.8% (39.2–61.2) | 59.8% (45.9–71.3) |

CR 60 57 54 47 40 36 32 26 7 7 7 5 4 4 4 3 2 1 1 0

1

4

PR 9 8 6 6 4 4 3 3 2 1 1 1 0

Nonresponder 14 7 5 4 2 2 2 2 0

^aReverse Kaplan-Meier method was used to obtain median follow-up and its 95% CI; ^bKaplan-Meier method was used to obtain 2-sided 95% CI intervals.

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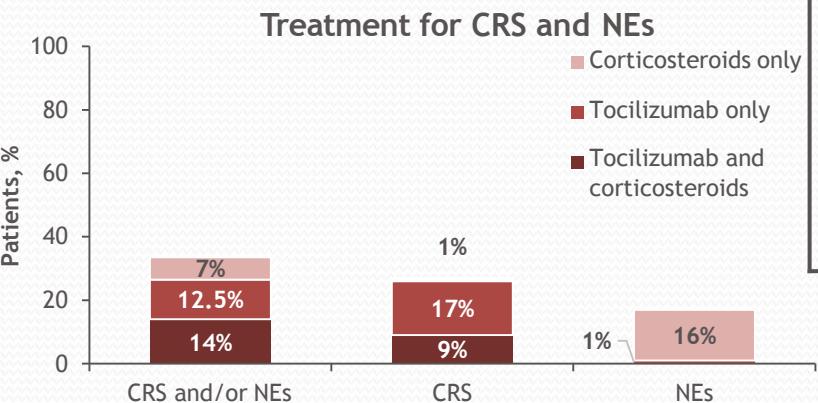
Wang M, et al. ICML 2023 [Abstract #LBA3]

Treatment-emergent AESIs and management of CRS and NEs

| Other AESIs | Liso-cel–treated set (n = 88) |
|---|----------------------------------|
| Prolonged cytopenias, ^c n (%) | 35 (40) |
| Grade ≥ 3 infections, ^d n (%) | 13 (15) |
| Hypogammaglobulinemia, n (%) | 6 (7) |

| Patients with CRS and NEs | Liso-cel–treated set (n = 88) |
|---|-------------------------------|
| CRS, ^a n (%) | |
| Any grade | 54 (61) |
| Grade 1/2 | 53 (60) |
| Grade 3 | 0 |
| Grade 4 | 1 (1) |
| Grade 5 | 0 |
| Median (range) time to onset, days | 4.0 (1–10) |
| Median (range) time to resolution, days | 4.0 (1–14) |
| NEs, ^b n (%) | |
| Any grade | 27 (31) |
| Grade 1/2 | 19 (22) |
| Grade 3 | 7 (8) |
| Grade 4 | 1 (1) |
| Grade 5 | 0 |
| Median (range) time to onset, days | 8.0 (1–25) |
| Median (range) time to resolution, days | 5.0 (1–45) |

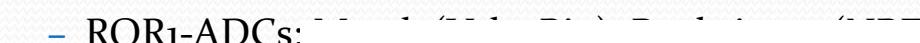
Treatment for CRS and NEs



^aCRS was graded based on the Lee 2014 criteria; ^bNEs were defined as investigator-identified neurological AEs related to liso-cel; ^cDefined as grade ≥ 3 laboratory abnormalities of neutropenia, anemia, and/or thrombocytopenia at Day 30 after liso-cel infusion; ^dIncludes grade ≥ 3 TEAEs from the infections and infestations (system organ class) by AE high-level group term. AESI, adverse event of special interest; NE, neurological event.

ROR1 (Receptor Tyrosine Kinase-Like Orphan Receptor 1)

Compelling Tumor-Specific Target

- Expressed on **most B-cell malignancies**, including
 - Mantle cell lymphoma (MCL), diffuse large B-cell lymphoma (DLBCL), chronic lymphocytic leukemia (CLL)
- Expressed on **many solid tumors**
 - Increased ROR1 expression associated with more aggressive tumors, shorter PFS and OS
- ROR1 activity associated with **aggressive phenotype**
 - Invasion, metastasis, stem cell-like behavior, and resistance to treatment
- Subject of **large pharma acquisitions**
 - ROR1-ADCs: 
- **Oncternal ROR1 pipeline differentiated and advancing**
 - Deep target biology expertise & immunotherapy experience



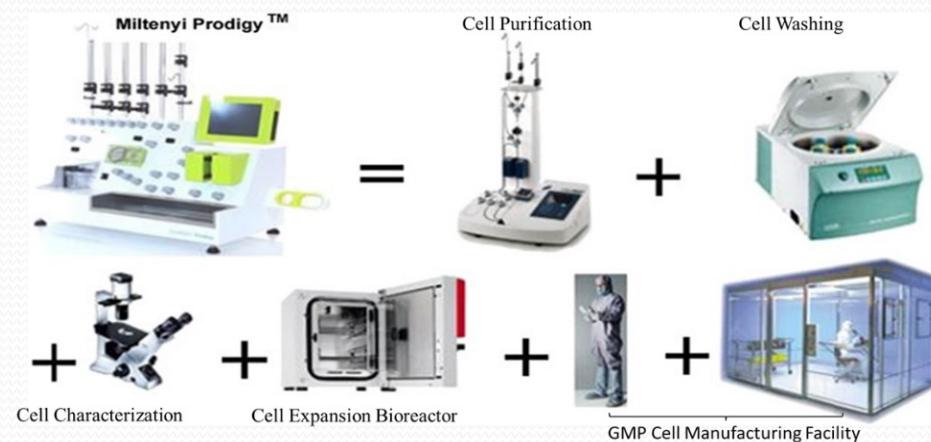
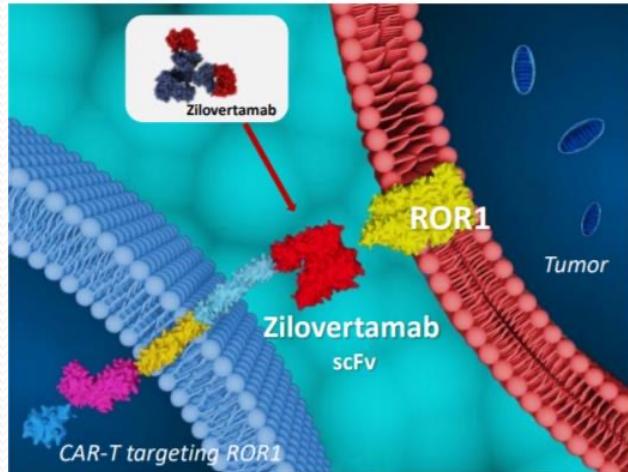
ROR1 Expressed on Multiple Solid and Liquid Tumors

| | |
|------------|-----|
| MCL | 95% |
| CLL | 95% |
| Uterus | 96% |
| Lymphoma | 90% |
| Prostate | 90% |
| Skin | 89% |
| Pancreatic | 83% |
| Adrenal | 83% |
| Lung | 77% |
| Breast | 75% |
| Testicular | 73% |
| Colon | 57% |
| Ovarian | 54% |

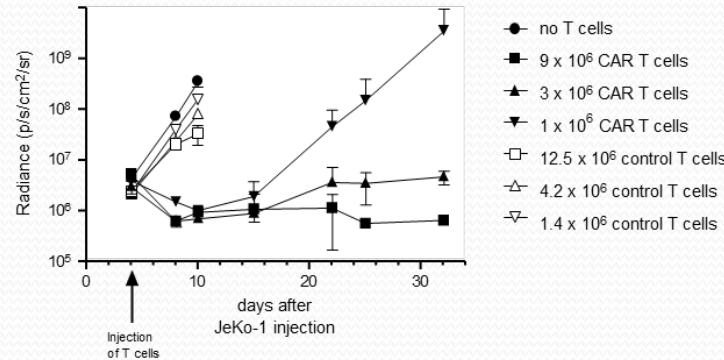
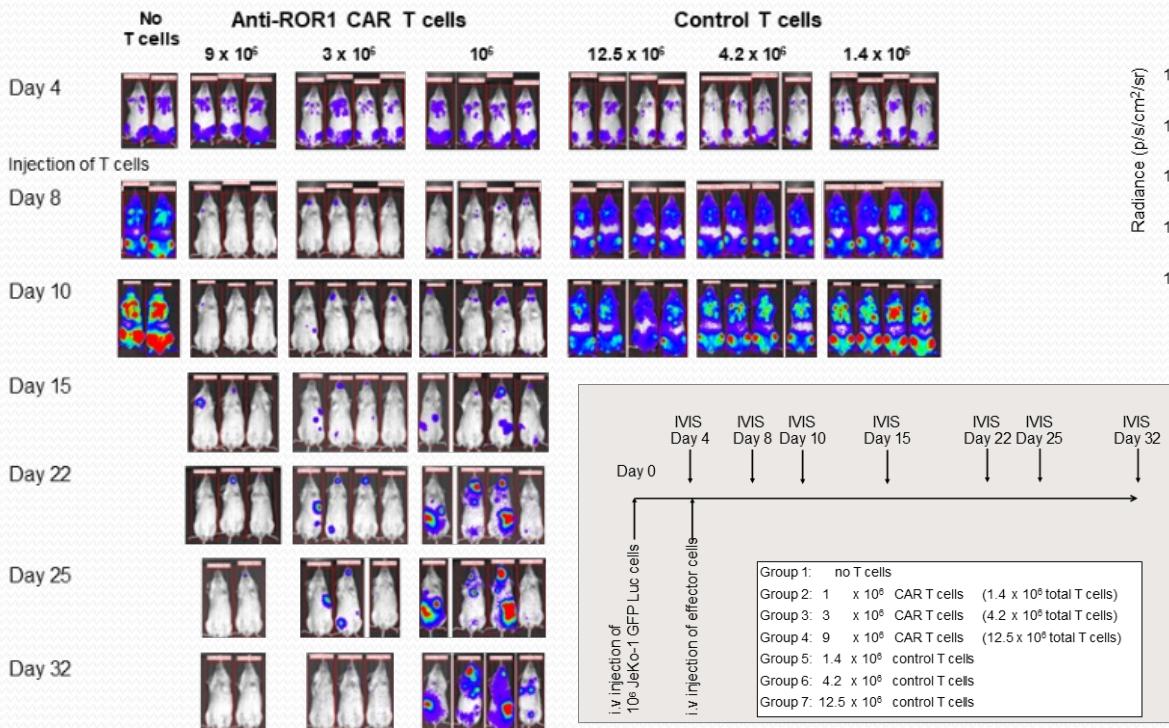
Zhang 2012
AJP

ONCT-808 – CMC and Manufacturing

1. Lead ROR1 CAR construct optimized and selected with demonstrated high potency against ROR1+ cancer cell lines
2. Lentivirus production process confirmed
3. Oncternal ROR1 CAR-T cell product process optimized and confirmed
 - Leveraging a flexible, closed fully-automated platform
 - 8-day production process post-activation
 - Greater than 2 billion CAR+ T cells produced with over 60% CAR+ expression
 - Majority of CAR T cells with juvenile phenotypes (CD4 and CD8 stem central memory T cells)
4. Harvard/Dana Farber CMCF (Cell Manipulation Core Facility) agreed for Phase 1 manufacturing



ONCT-808 – Strong Anti-tumor Activity in Preclinical Xenograft Model



Data generated in collaboration with
Dr. Evren Alici (Karolinska Institutet).

Data were presented at EHA 2022.

- Strong anti-tumor activity of ROR1 CAR-T cells demonstrated in MCL xenograft mouse model
- Data from additional IND-supporting in vivo studies will be presented at upcoming scientific conferences

MCL: Single Center (MCW) Study Using Anti- CD20/Anti-CD19 CAR

Original Reports | Hematologic Malignancy



Phase I/II Study of Adaptive Manufactured Lentiviral Anti-CD20/Anti-CD19 Chimeric Antigen Receptor T Cells for Relapsed, Refractory Mantle Cell Lymphoma

Nirav N. Shah, MD, MS¹ ; Alfredo S. Colina, BA² ; Bryon D. Johnson, PhD¹; Aniko Szabo, PhD³; Fateeha Furqan, MD¹; Tyce Kearn, MD, PhD¹ ; Dina Schneider, PhD⁴ ; Marleny Vargas-Cortes, BS⁵ ; Jessica L. Schmeling, BS⁵ ; Michael B. Dwinell, PhD²; Katie Palen, BS¹; Walter Longo, MD¹; Peiman Hematti, MD¹ ; Anthony E. Zamora, PhD² ; Parameswaran Hari, MD, MS¹ ; Daniel Bucklan, MD⁶; Ashley Cunningham, MD⁷; Mehdi Hamadani, MD¹ ; and Timothy S. Fenske, MD¹ 

DOI <https://doi.org/10.1200/JCO-24-02158>

ASCO® Journal of Clinical Oncology®

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MCL: Patient Characteristics

TABLE 1. Patients With MCL (N = 17)

| Baseline Patient Characteristic | Phase I = 3, Phase II = 14 |
|---|----------------------------|
| Age, years, median (range) | 63 (50-74) |
| Male sex, No. (%) | 15 (88) |
| Previous auto-HCT, No. (%) | 8 (47) |
| Previous allo-HCT, No. (%) | 2 (12) |
| LDH >normal on day 0, No. (%) | 6 (35) |
| Marrow involvement before CAR infusion, No. (%) | 14 (82) |
| BTKi exposed, No. (%) | 16 (94) |
| BTKi progressed, No. (%) | 13 (76) |
| Noncovalent (pirtobrutinib) BTKi progressed, No. (%) | 6 (35) |
| Previous lines (including transplant), median (range) | 4 (2-8) |
| Previous bendamustine, No. (%) | 13 (76) |
| Previous bendamustine <1 year, No. (%) | 2 (12) |
| MIPI at diagnosis (n = 14), No. (%) | |
| Low | 6 (35) |
| Intermediate | 4 (31) |
| High | 4 (31) |
| Missing | 3 (18) |
| Complex cytogenetics, No. (%) | 3 (18) |
| p53 aberrations, No. (%) | 8 (47) |
| p53 mutation | 6 (35) |
| 17p deletion by FISH or cytogenetics | 3 (18) |

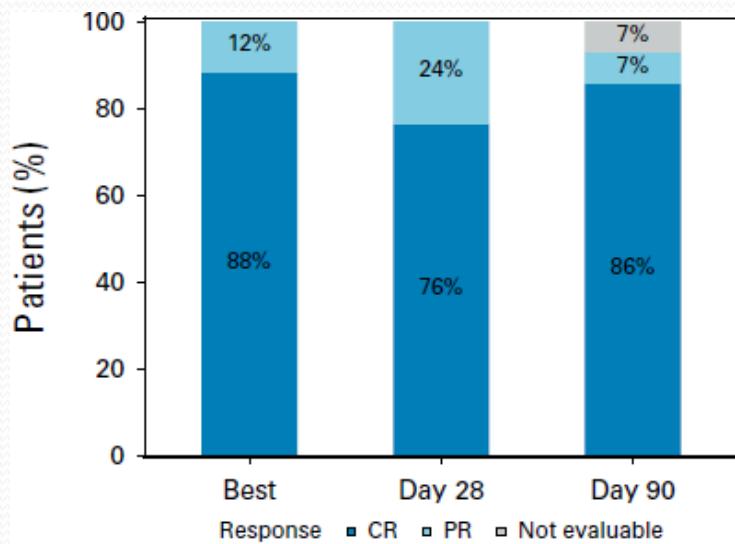
Supplemental Table 1: p53 Aberrations and Complex Cytogenetics by Subject

| Subject Number | P53 aberration | Karyotype |
|----------------|---|----------------------|
| 3 | Loss of chromosome 17 | Complex Karyotype |
| 5 | N/A | N/A |
| 7 | Positive for TP53 mutation | N/A |
| 12 | N/A | -Y/Not Complex |
| 15 | Negative for TP53 mutation | Not Complex |
| 16 | Negative for TP53 mutation | N/A |
| 19 | Negative for TP53 mutation | Not Complex |
| 20 | TP53 deletion by FISH Positive for TP53 mutation | Not Complex |
| 30 | N/A | Complex karyotype |
| 31 | TP53 deletion by FISH | Complex karyotype |
| 40 | Positive for TP53 mutation | -Y/Not Complex |
| 42 | Negative for TP53 mutation | Del(13q)/Not Complex |
| 43 | N/A | Not Complex |
| 49 | Positive for TP53 mutation | Not Complex |
| 55 | N/A | Not Complex |
| 56 | Positive for TP53 mutation | N/A |
| 57 | Positive for TP53 mutation | Not Complex |

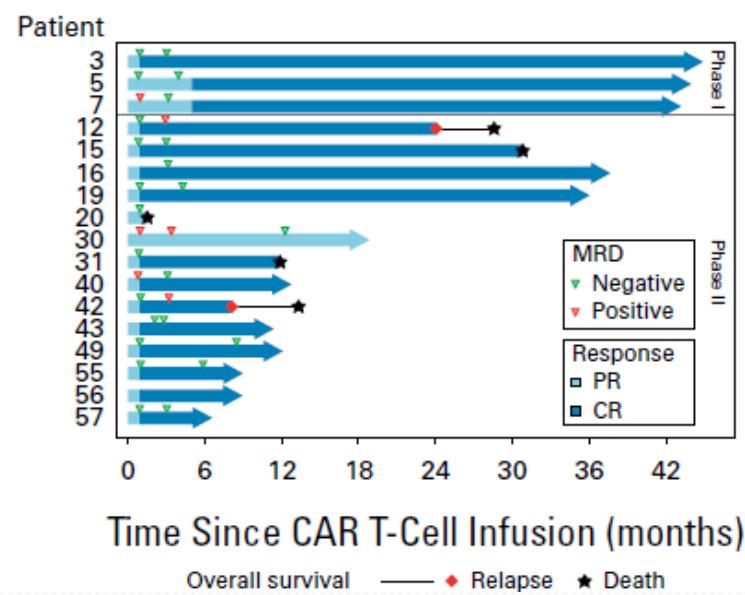
N/A- Not available or unable to assess

MCL: Clinical outcomes for patients treated with LV20.19 CAR T cells

Percentages of the 17 patients who had clinical response at day 28, day 90, and best overall response



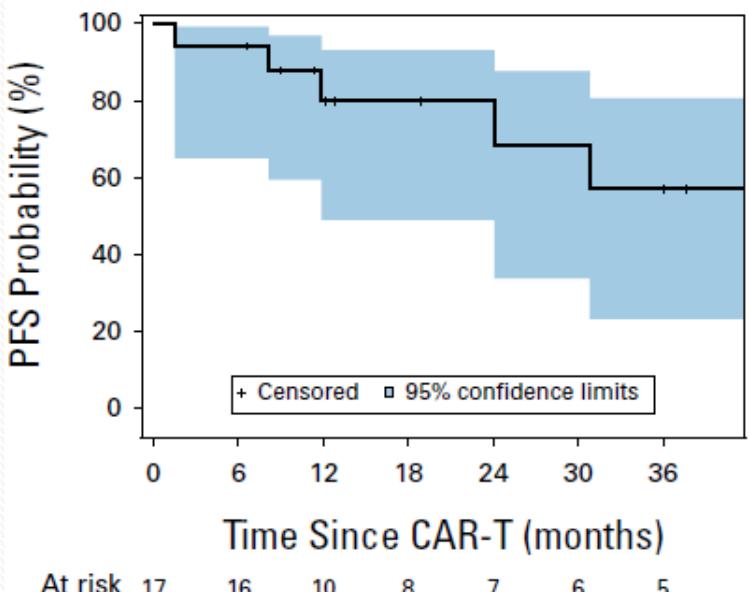
Patient identification on the y-axis and demarcation for MRD status and response



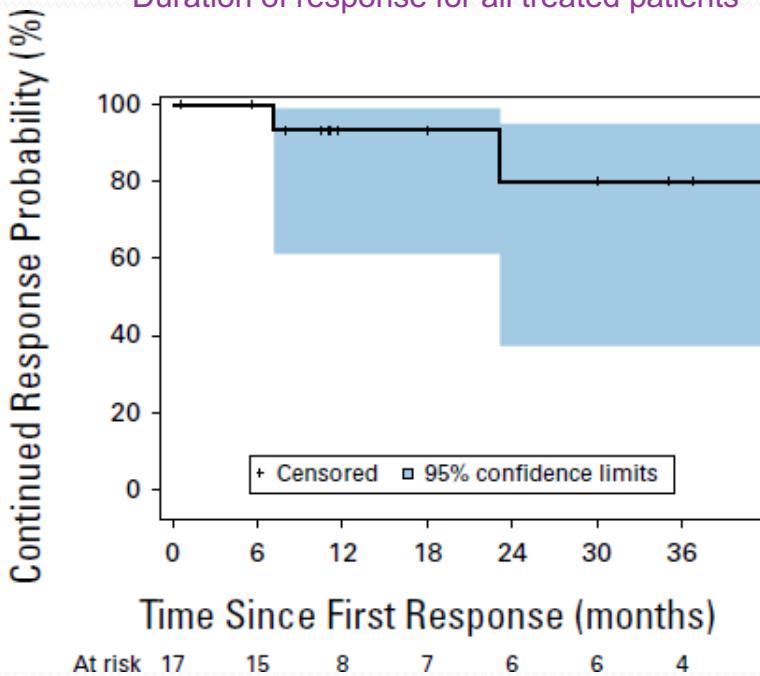
Shah NN, Colina AS, Johnson BD, et al. Phase I/II Study of Adaptive Manufactured Lentiviral Anti-CD20/Anti-CD19 Chimeric Antigen Receptor T Cells for Relapsed, Refractory Mantle Cell Lymphoma. *Journal of Clinical Oncology*. Published online March 31, 2025. Accessed April 3, 2025. <https://ascopubs.org/doi/10.1200/JCO-24-02158>

MCL: Clinical outcomes for patients treated with LV20.19 CAR T cells

PFS of all treated patients

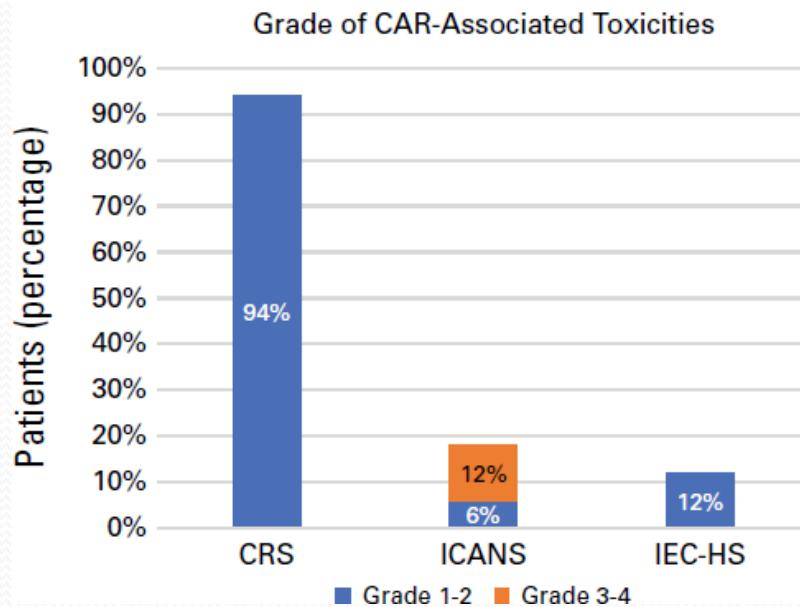


Duration of response for all treated patients

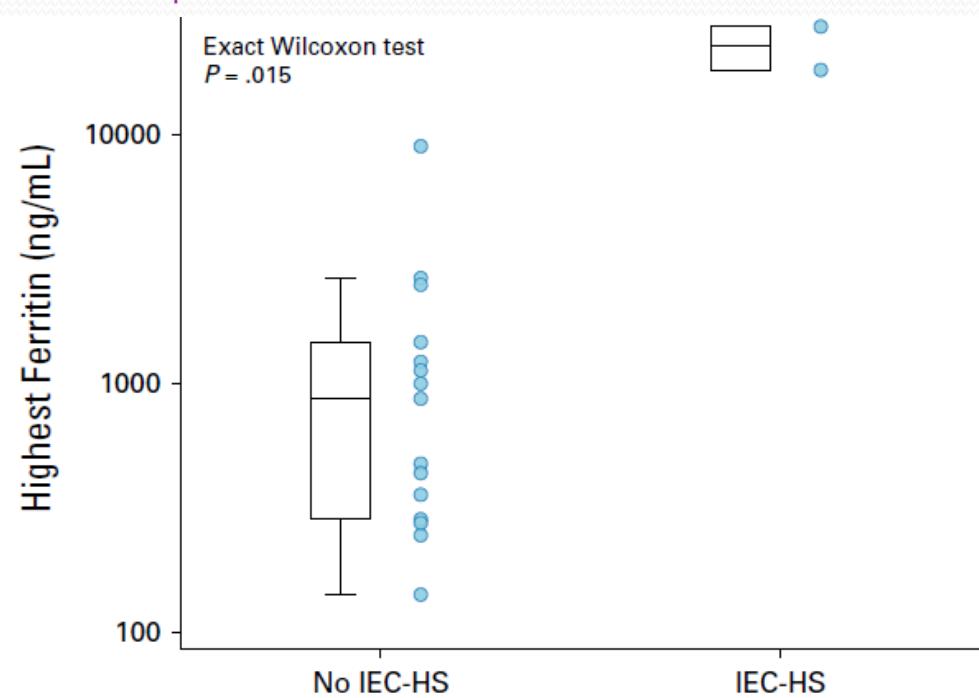


MCL: Safety data for patients treated with LV20.19 CAR T cells

Percentages of patients (n=17) who experienced CRS, ICANS, or IEC-HS



Peak ferritin levels are depicted on the basis of presence or absence of IEC-HS



Shah NN, Colina AS, Johnson BD, et al. Phase I/II Study of Adaptive Manufactured Lentiviral Anti-CD20/Anti-CD19 Chimeric Antigen Receptor T Cells for Relapsed, Refractory Mantle Cell Lymphoma. *Journal of Clinical Oncology*. Published online March 31, 2025. Accessed April 3, 2025. <https://ascopubs.org/doi/10.1200/JCO-24-02158>



BAFFR CAR T Cells (PMB-CT01) Demonstrate Durable Responses and Manageable Toxicities in Relapsed/Refractory B-Cell Lymphomas with Prior CD19-Directed Therapy Failure or CD19-Negative Disease

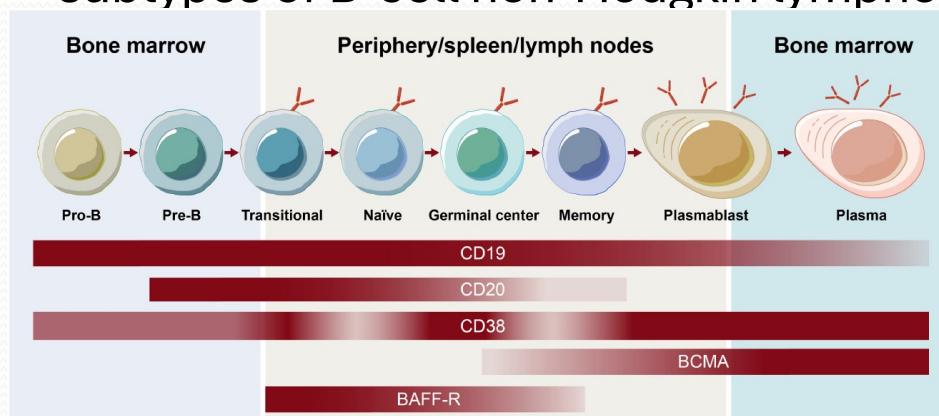
L. Elizabeth. Budde, Marissa M. Del Real, John H. Baird, Lu Chen, Joo Y. Song, Xiuli Wang, Swetha Thiruvengadam, Marie Hu, Alan Macias, Emanuela Marcucci, Soungchul Cha, Zhenyuan Dong, Teresa Kim, Baishakhi Barva, Sandrine Puverel, Qing Liu-Michael, Hazel (Ting-Ying) Cheng, Stephen J. Forman, and Larry W. Kwak

City of Hope National Medical Center
Duarte, CA, USA

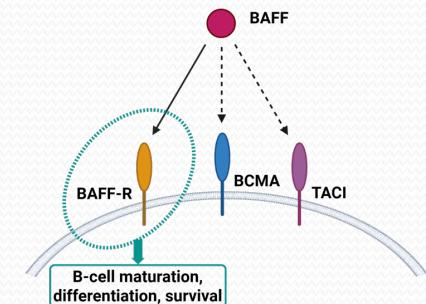


BAFF-R as a novel therapeutic target in B-cell lymphomas

- BAFF-R (B-cell activating factor receptor) is a member of the TNF superfamily and the main receptor for BAFF.
- It is selectively expressed on B cells and on most subtypes of B-cell non-Hodgkin lymphomas



Robinson et al., Front Immunol. 2024;15:1454747



| B-cell malignancy | BAFF-R positive cases (%) |
|-------------------------------|---------------------------|
| Hairy cell leukemia | 10/10 (100) |
| Chronic lymphocytic leukemia | 21/21 (100) |
| Mantle cell lymphoma | 7/7 (100) |
| Follicular lymphoma | 13/16 (81) |
| Diffuse large B-cell lymphoma | 14/18 (78) |
| Marginal zone lymphoma | 10/11 (91) |

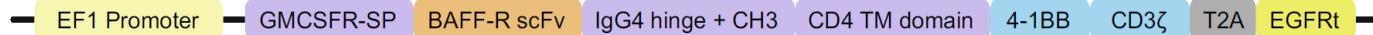
Rodig et al., Hum Pathol. 2005;36(10):1113-9

BAFF-R as a novel therapeutic target in B-cell lymphomas

- BAFF-R signaling promotes normal B-cell proliferation and is required for survival
- In lymphoid malignancy, BAFF-R signaling activates NF-κB pathways and contribute to malignant lymphoid cell survival and proliferation (ALL, MCL).
- This critical feature may limit the capacity of B-cell tumors to escape therapy by down-regulation of BAFF-R expression, as this would compromise their viability
- CD19-negative primary ALL tumors retained BAFF-R expression

Rodig *et al.*, *Hum Pathol.* 2005;36(10):1113-9; Novak *et al.*, *Blood* 2004;104(8):2247-53; Maia *et al.*, *PLoS One* 2011;6(6):e20787; Pham *et al.*, *Blood* 2011;117(1):200-10

- Kwak's group at COH generated a humanized BAFF-R Ab and incorporated the scFv into a CAR.

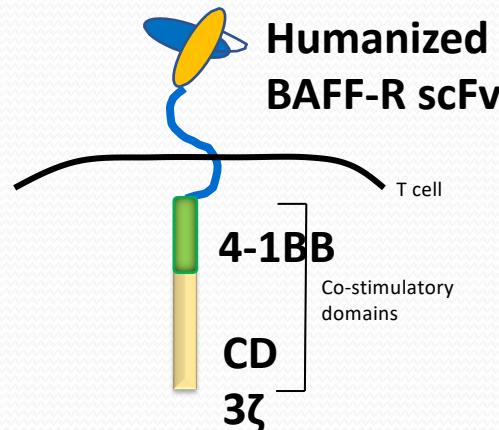


Published OnlineFirst November 27, 2017; DOI: 10.1158/1078-0432.CCR-17-1193

Cancer Therapy: Preclinical

Novel BAFF-Receptor Antibody to Natively Folded Recombinant Protein Eliminates Drug-Resistant Human B-cell Malignancies *In Vivo*

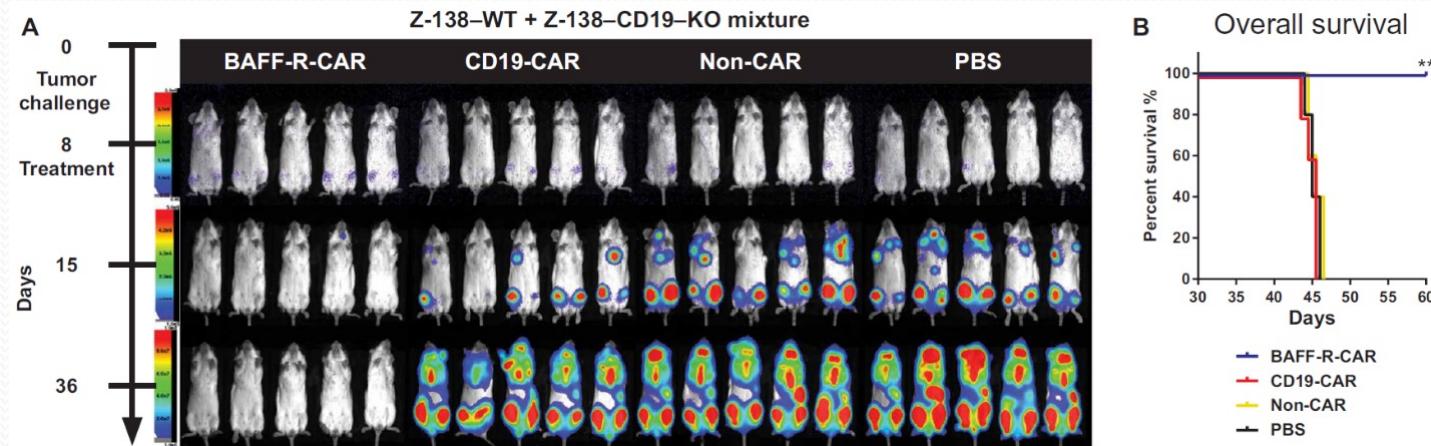
Hong Qin¹, Guowei Wei¹, Ipppei Sakamaki², Zhenyuan Dong¹, Wesley A. Cheng¹, D. Lynne Smith¹, Feng Wen^{1,3}, Han Sun¹, Kunhwa Kim⁴, Soungchul Cha⁴, Laura Bover⁵, Sattva S. Neelapu⁴, and Larry W. Kwak¹



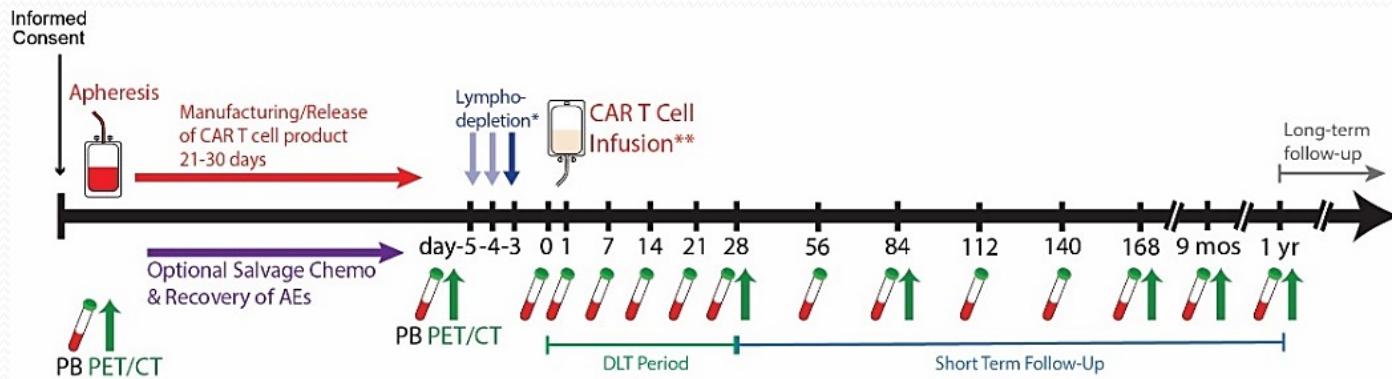
- Bench to Bedside Development of a Novel, Personalized Cellular Therapy for Blood Cancers

Preclinical studies support BAFF-R as a B-NHL target

- Head-to-head comparisons with CD19 CAR T cells in preclinical models suggest superior efficacy with BAFF-R CAR T cells
- BAFF-R CAR T cells can eliminate lymphoma cells regardless of CD19 expression



First-in-human, multicenter phase 1 trial (NCT05370430)



| Dose level | Dose of BAFFR-CAR T Cells* |
|--------------------------|--|
| -1 (de-escalation) | 20×10^6 cells |
| 1 (starting dose) | 50×10^6 cells |
| 2 | 200×10^6 cells |
| 3 | 600×10^6 cells |

* ≤20% lower cell dose permitted

- Dose-finding cohort (3+3) (**completed**)
- Three histology expansion cohorts (12 pts each) (MCL, LBCL, FL) at RP2D

6 sites: City of Hope Duarte, Stanford University, University of Minnesota, Atrium Health Levine Cancer Institute, Providence Swedish Cancer Institute, University of Kansas Cancer Center

Eligibility and Study objectives

Inclusion

- BAFF-R+ B-NHL
- ≥ 18 years old
- Measurable disease
- ECOG ≤ 2
- Prior CAR T allowed, ≥ 90 days from leukapheresis

Exclusion

- Active CNS involvement
- Prior allo-HCT
- Auto-HCT within 6 months
- Steroids and immunosuppressants

Primary objectives

- Safety
- MTD/RP2D

Secondary objectives

- Response
- Duration of B-cell aplasia
- PFS, OS

Exploratory objectives

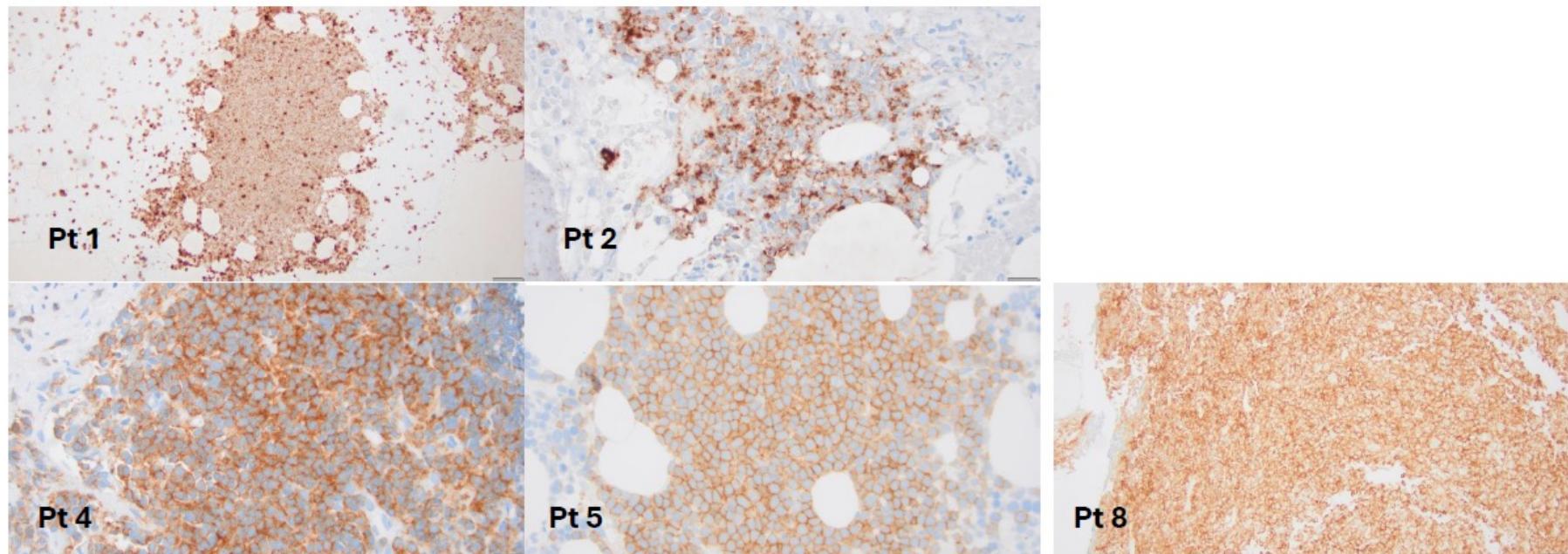
- Expansion, persistence
- MRD-negative rate
- Cytokines
- BAFF-R expression post-relapse or progression
- CAR T polyfunctionality

Baseline characteristics

- Nine patients were infused

| Characteristic | Dose Level 1 | | | Dose Level 2 | | | | | |
|---------------------|--------------|------------|---------------|--------------|------------|------------|------------|------------|------------|
| | Patient 1 | Patient 2 | Patient 3 | Patient 4 | Patient 5 | Patient 6 | Patient 7 | Patient 8 | Patient 9 |
| Gender | M | M | M | M | M | M | F | M | M |
| Age | 56 | 75 | 41 | 62 | 72 | 74 | 62 | 57 | 58 |
| Histology | MCL | MCL | THRBCL | MCL | MCL | MZL | FL | MCL | MCL |
| • Stage at baseline | IV | IV | III | IV | IV | IIA | III | IV | IV |
| • # Prior lines | 4 | 10 | 3 | 3 | 3 | 1 | 7 | 3 | 6 |
| • Prior CD19 CAR T | Yes | Yes | No | No | Yes | No | Yes | Yes | Yes |
| • Prior HCT | No | No | Yes | No | No | No | No | Yes | No |
| • Prior TCE | No | Yes | No | Yes | No | No | Yes | No | No |
| • CD19 expression | Yes | Yes | No | Yes | Yes | Yes | Yes | Yes | Yes |
| • CD20 expression | Yes | Yes | No | Yes | Yes | Yes | No | Yes | Yes |
| • TP53 mutation | Yes | No | No | No | Yes | No | No | Yes | n/a |

BAFF-R Expression



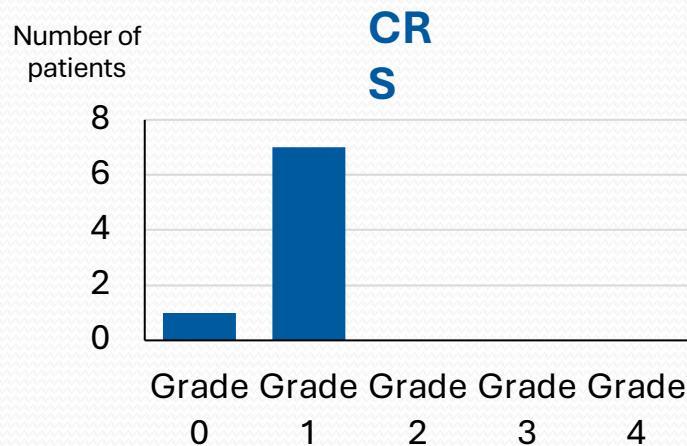
Treatment safety

CTCAE grade ≥ 3 AE considered at least possibly related to the BAFF-R CAR T cells

| No DLTs | | Grade | |
|---------|---|---------|---------|
| | | Grade 3 | Grade 4 |
| | Cardiac disorders | | |
| | Sinus tachycardia | 1 | 0 |
| | Infections and infestations | | |
| | Pneumonia viral | 1 | 0 |
| | Hematologic events | | |
| | Lymphocyte count decreased | 3 | 4 |
| | Neutrophil count decreased | 1 | 2 |
| | Platelet count decreased | 1 | 2 |
| | White blood cell count decreased | 4 | 0 |
| | Metabolism and nutrition disorders | | |
| | Hypophosphatemia | 1 | 0 |
| | Skin and subcutaneous tissue disorders | | |
| | Rash | 1 | 0 |

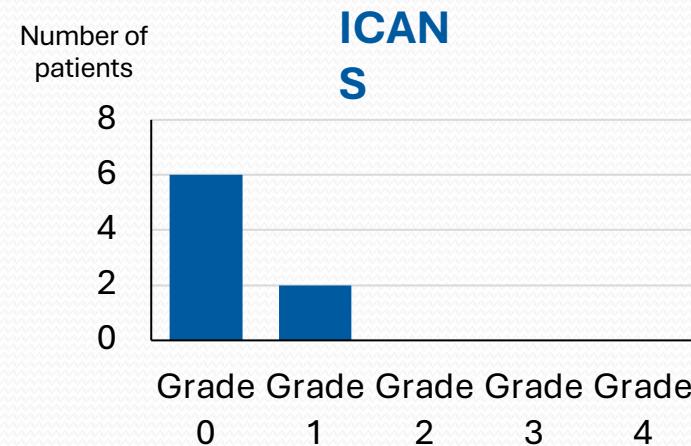
One patient developed a myelodysplastic syndrome that was deemed unrelated to study treatment

Treatment safety



87.5% grade 1 CRS (7/8 patients)

No grade >1 CRS

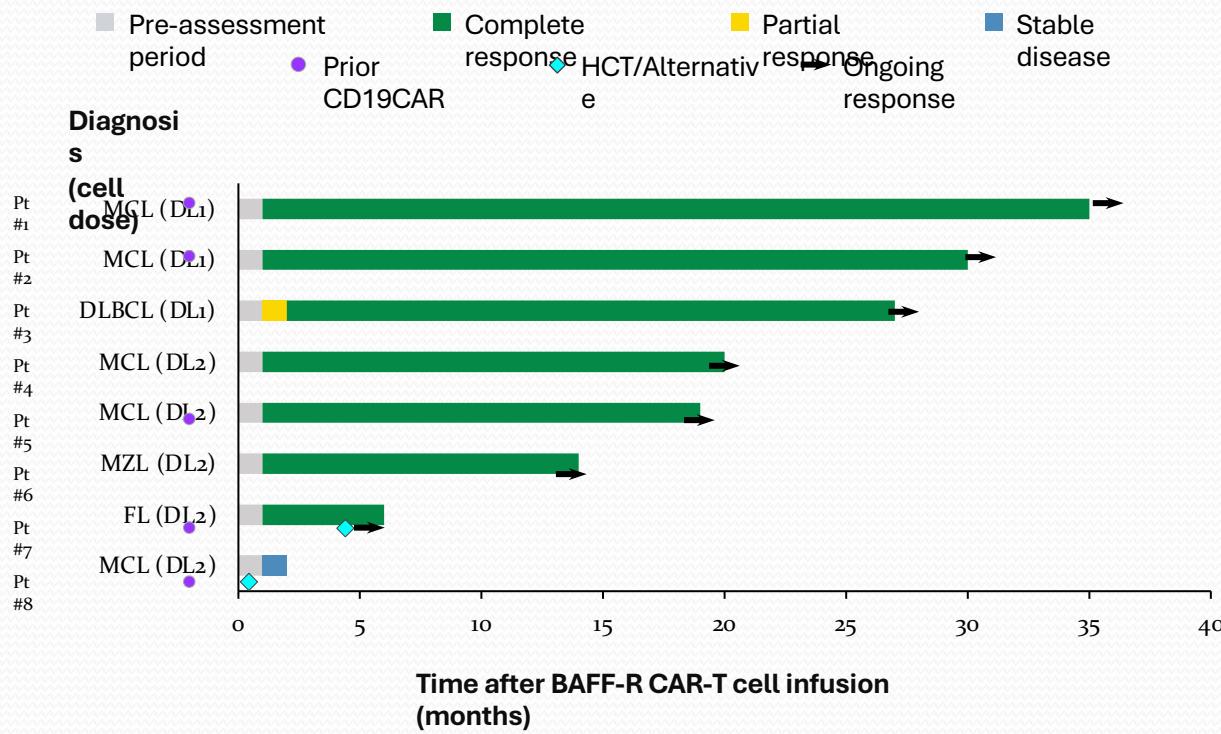


25% grade 1 ICANS (2/8 patients) that resolved without corticosteroids

No grade >1 ICANS

67% of patients (6/9) received LD and CAR T infusion in the outpatient setting

Response to treatment

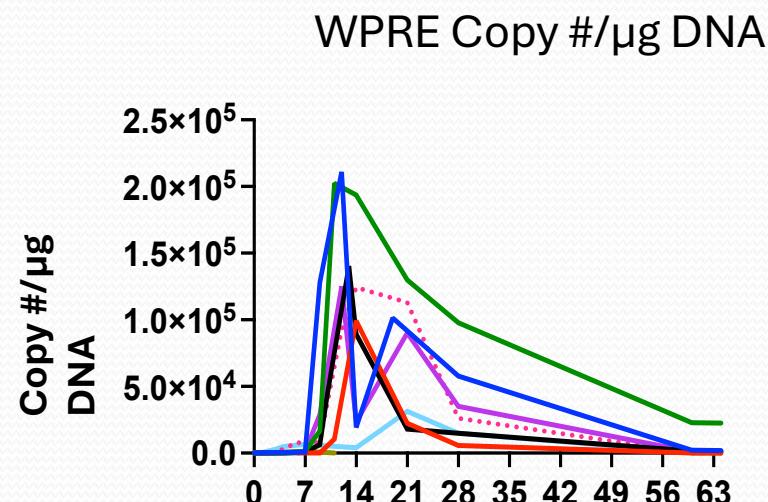
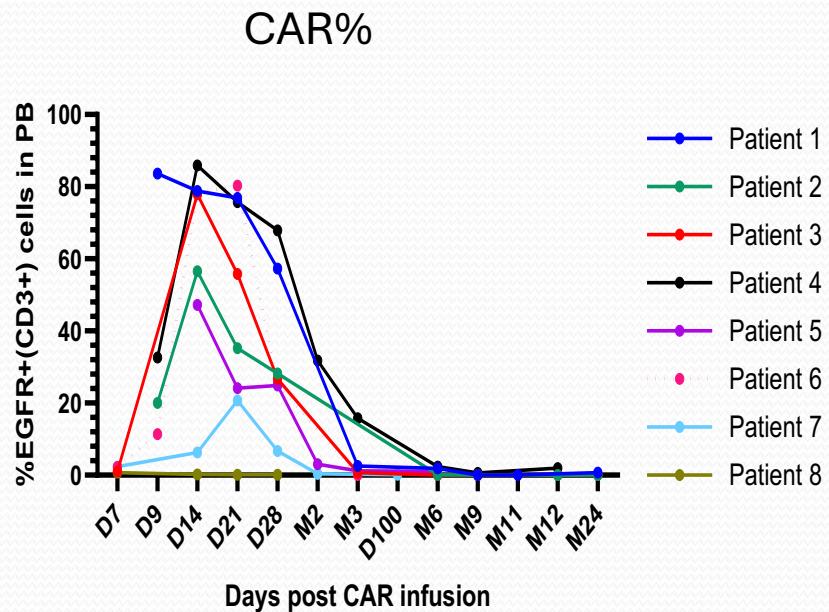


87.5% CR at 3 months
(7/8 patients)

All 4 MCL CR patients
were MRD-negative by
flow and NGS

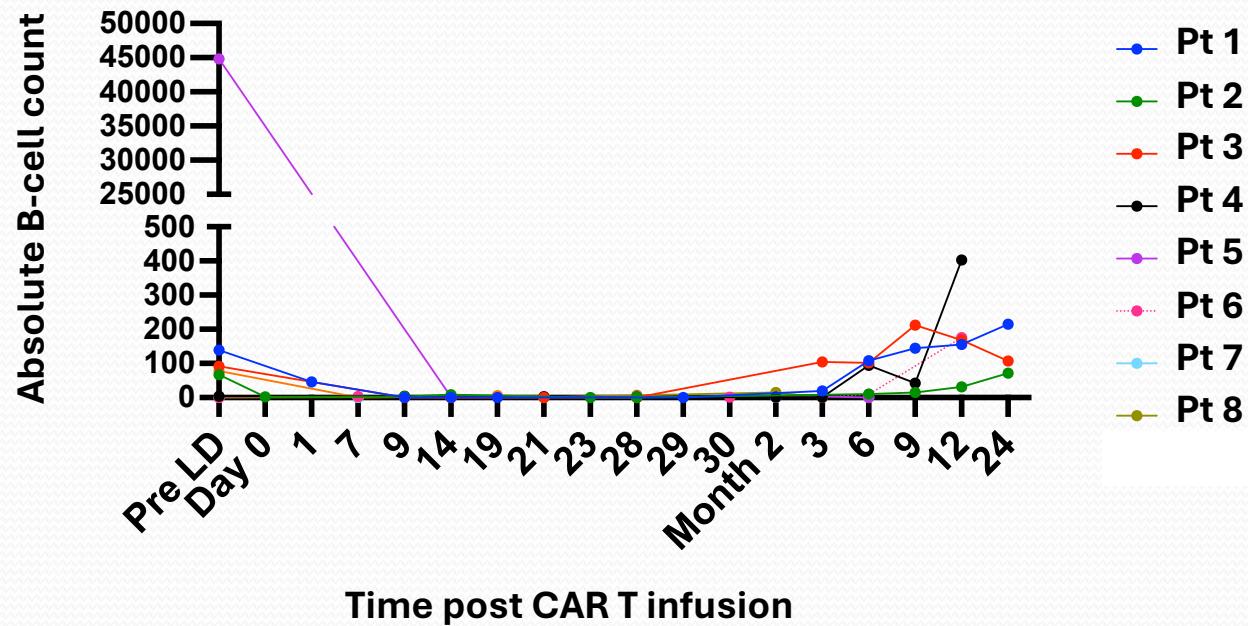
No relapses

Robust BAFF-R CAR T expansion



- **Robust CAR-T cell expansion** was observed in all responders with peak of expansion on day 12-21

B-cell recovery after BAFF-R CAR T cell infusion





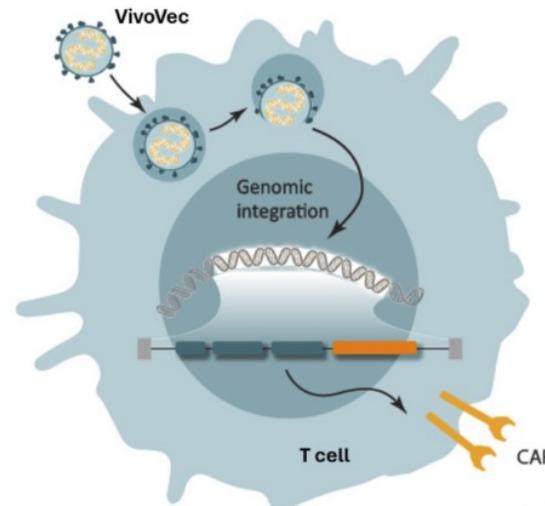
Advances in In Vivo CAR T-Cell Therapy

01/15/2026

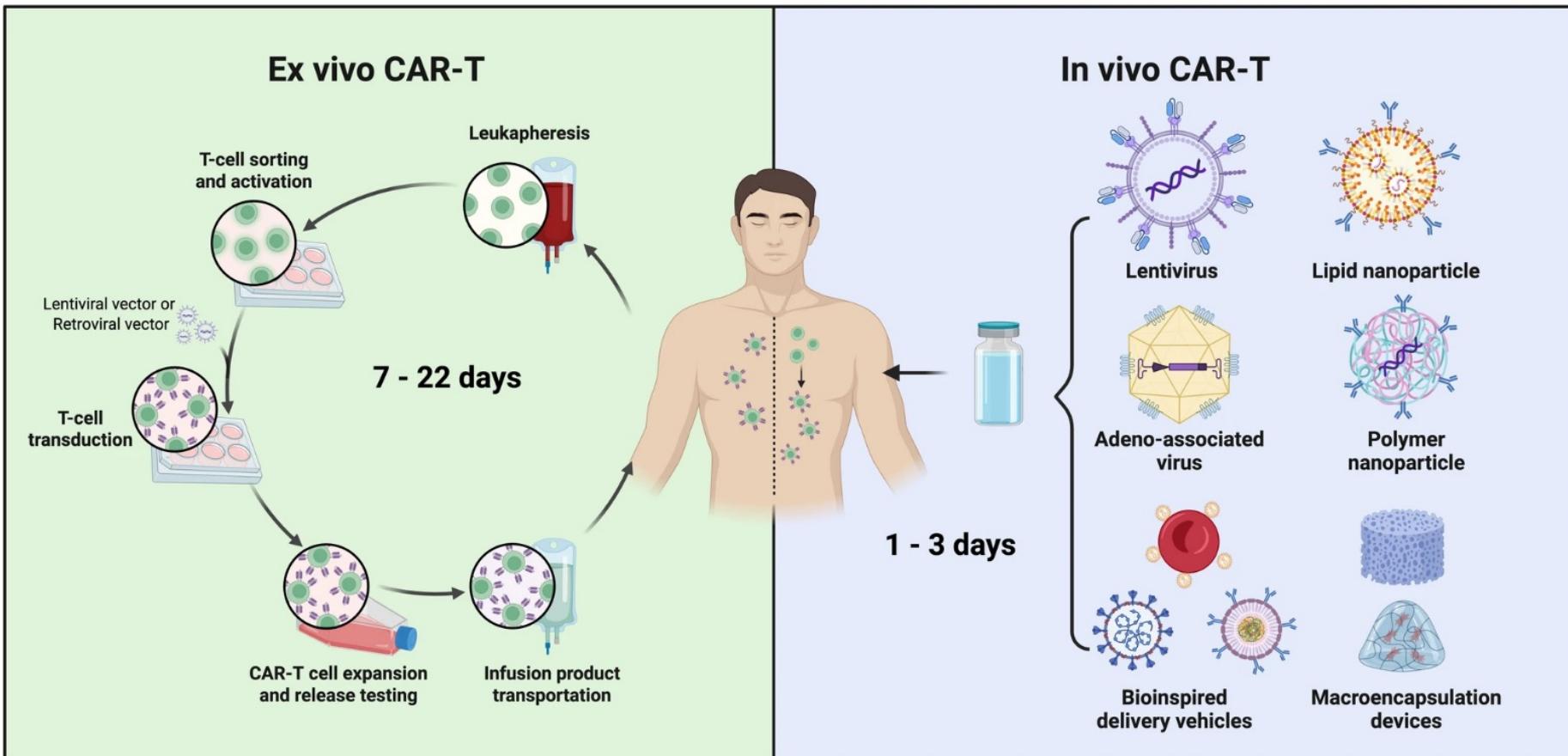
In Vivo CAR-T Therapy

- In vivo CAR-T therapy is an emerging approach in which CAR constructs are delivered directly into patients, enabling T cells to be engineered *in situ* without ex vivo manufacturing.
- Using platforms such as engineered viral vectors or targeted nanoparticles, this strategy aims to improve accessibility, reduce cost, and accelerate treatment, while introducing distinct challenges in delivery and safety.

In vivo CAR T cell therapy =
T cells are genetically altered *within* the body
to enhance their effectiveness at fighting
against disease (eg, cancer); drug product
is available immediately to patients instead
of within weeks or months.^{19,20}



From Multi-Step Ex Vivo Manufacturing to Streamlined In Vivo CAR-T Therapy



Lentiviral-based *in vivo* CAR-T cell platforms in development

| Company | Targeting mechanism | Therapeutic payloads | Lead indications | Preclinical evidence | Development stage |
|-------------------|--|---|---|---|---|
| Juno Therapeutics | Anti-CD7 scFv-decorated particles (T cell and natural killer cell engineering) | Anti-CD20 CAR, anti-CD19 CAR | B cell malignancies, autoimmunity | Proof of principle in mice and NHPs ⁸⁴ | Clinical (phase I enrolling in 2024 with anti-CD20 CAR) ^{78,85} |
| | Multi-domain anti-CD3, CD80, CD58 decorated particles (T cell engineering) | Anti-CD19 CAR, anti-CD22 CAR, anti-CD20 CAR | B cell malignancies, autoimmunity | Proof of principle in mice and NHPs ^{81,88,91} | Clinical (phase I initiated in 2024 with anti-CD19 CAR, the others in 2024, 2026) ⁹⁰ |
| | Anti-CD3 decorated particles | Anti-CD19 CAR | B cell malignancies | Not disclosed | Investigator-sponsored trial, first responder in a patient with lymphoma ^{95,96} |
| | Targeted lentiviral particles | Anti-BCMA CAR, undisclosed | Multiple myeloma, autoimmunity, solid tumours | Proof of principle in mouse model ⁹⁵ | Phase I initiation in 2025, first clinical response in myeloma, acquisition ^{98,101,148} |
| | Anti-CD3 decorated particles | Anti-BCMA CAR | Multiple myeloma | Proof of principle in mice and NHPs ^{102,103} | Phase I initiation mid 2025 |
| | Anti-CD8 fusogen-decorated particles | Anti-CD19 CAR | Undisclosed | Proof of principle in mice and NHPs ^{78,109,111} | Undisclosed |
| | CD46-targeted viral-like particles (multilineage) | Anti-HER2 CAR | Multiple solid tumours | Proof of principle in preclinical models ¹¹² | Undisclosed |
| | CD3-targeted lentiviral vector | Anti-CD19 CAR | B cell malignancies | Proof of principle, mouse models ¹¹⁴ | Undisclosed |

BCMA, B cell maturation antigen; CAR, chimeric antigen receptor; NHP, non-human primate.

LNP-RNA-based *in vivo* CAR-T cell platforms in development

| Company | Targeting mechanism | Therapeutic payloads | Lead indications | Preclinical evidence | Development stage |
|----------------------|---|---|---|---|---|
| LNP-RNA Therapeutics | LNPs – macrophage tropic | Anti-Trop2 CAR, anti-GPC3 CAR, anti-HER2 CAR, anti-gp75 CAR (CD89 and natural killer cell p44-based CAR constructs) | Multiple solid tumours and hepatocellular carcinoma | Multiple preclinical models ^{122,124} | Clinical (phase I initiated in 2024 with anti-TROP2 and GPC3 CARs) and first clinical response ^{125,127} |
| | Anti-CD8 monoclonal antibody-decorated LNPs (CTL engineering) | Anti-CD19 CAR and undisclosed | Autoimmunity and undisclosed | Mouse and NHP proof of principle ¹³⁵⁻¹³⁷ | Phase I initiated (NCT06917742) |
| | tLNPs (T cells, myeloid cells and natural killer cells) | B cell-targeted CAR (RNA format) | CD19 ⁺ B cell malignancies | Undisclosed | Phase I initiated with first patient dosed ¹⁴⁰ |
| | CD8 T cell tLNPs | Anti-CD19 CAR (mRNA format) | Systemic lupus erythematosus | Mouse and NHP proof of principle ¹⁴² | Phase 1 initiated and evidence of activity reported ^{140,143} |
| | LNPs containing immunotropic lipids (pan-T cell engineering) | Anti-CD19 CAR (circular RNA format) | B cell malignancies and autoimmunity | Mouse and NHP proof of principle ¹⁴⁸ | Phase I initiation by 2026 |
| | tLNPs (T cell engineering) | Anti-CD22 and anti-CD19 CAR (mRNA) | Oncology, autoimmunity | Mouse and NHP proof of principle ^{152,154} | Undisclosed |
| | LNPs – macrophage tropic | Anti-GPC3 CAR and not disclosed | Hepatocellular carcinoma | Preclinical mouse models ^{161,162} | Preclinical stage; strategic changes announced ¹⁶⁴ |
| | tLNPs | Anti-CD19, CD20, BCMA using RNA writer (integrating payload) | Oncology, autoimmunity | Preclinical modelling ¹⁷¹⁻¹⁷³ | Undisclosed |
| | tLNPs and viral-like particles | CAR (circular RNA format) – details not disclosed | Undisclosed | Undisclosed | Undisclosed |

BCMA, B cell maturation antigen; CAR, chimeric antigen receptor; CTL, cytotoxic T cell lymphocyte; GPC3, glypican 3; LNP, lipid nanoparticle; NHP, non-human primate; tLNP, targeted LNP.

Clinical Trials

| Company | Product name | Targeting Mechanism and Payloads | Disease | ClinicalTrials.gov ID | Phase | Study Start | Collaborators and Investigators |
|----------------------|--------------|----------------------------------|--|-----------------------|---------|-------------|--|
| | UB-VV111 | CD3-Cocal-LV-CD19 CAR | R/R large B-cell lymphoma (LBCL) and chronic lymphocytic leukemia (CLL). | NCT06528301 | Phase 1 | 2024-11 | <ul style="list-style-type: none"> • City of Hope • The David and Etta Jonas Center for Cellular Therapy • Washington University School of Medicine/Siteman Cancer Center • University of Nebraska Medical Center • University of Cincinnati Medical Center • Fred Hutch Cancer Center • Royal North Shore Hospital • St. Vincent's Hospital Melbourne |
| Myeloid Therapeutics | MT-303 | LNP-GPC3 scFv/CD89 (mRNA) | Advanced or Metastatic GPC3-Expressing Cancers, Including HCC | NCT06478693 | Phase 1 | 2024-07-01 | <ul style="list-style-type: none"> • CREATE Medicines • Australia (4 locations) • South Korea (3 locations) • Taiwan (2 locations) <p>https://clinicaltrials.gov/</p> |

First-in-Human Validation of Viral In Vivo CAR-T: ESO-T01



The Lancet

Volume 406, Issue 10500, 19–25 July 2025, Pages 228-231



Key Findings

- **Patients treated:** 4
- **Dose:** single dose, 2.0×10^8 TU
- **Overall response rate (ORR):** 100%
 - 2 CR, 2 PR
- **Extramedullary lesion clearance observed**
 - Indicates effective CAR-T infiltration into the TME
- **Acceptable safety profile**
 - No adverse events > Grade 3 (except hematologic toxicities)
- **CAR-T cellular kinetics**
 - Comparable to commercial ex vivo CAR-T products

Correspondence

In-vivo B-cell maturation antigen CAR T-cell therapy for relapsed or refractory multiple myeloma

Jia Xu ^a, Lin Liu ^a, Philippe Parone ^b, Wei Xie ^a, Chunyan Sun ^a, Zhaozhao Chen ^a, Jishuai Zhang ^c, Chunrui Li ^d, Yu Hu ^a, Heng Mei ^a 

^a Institute of Hematology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430022, China

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^d Department of Hematology, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

Xu J, et al. *Lancet*. 2025;406(10500):228-231.

Minimal residual disease (MRD)-negative outcomes following a novel, *in vivo* gene therapy generating anti-B-cell maturation antigen (BCMA) chimeric antigen receptor (CAR)-T cells in patients with relapsed and refractory multiple myeloma (RRMM): Preliminary results from inMMMyCAR, the first-in-human phase 1 study of KLN-1010

Simon Harrison¹, Phoebe Joy Ho^{2,3}, Sueh-Li Lim^{4,5}, Stephanie Talam⁶, Hannah Pahl⁷, Dharmesh Dingar⁸, Scott Currence⁸, Travis Quigley⁸, Andrew Spencer⁴

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⁵ Alfred Hospital, Malignant Haematology, Transplantation and Cellular Therapies Service, Melbourne, Australia

⁶ Royal Prince Alfred Hospital, Institute of Haematology, Camperdown, Australia

⁷ Peter MacCallum Cancer Centre, Parkville Cancer Clinical Trials Unit, Melbourne, Australia

⁸ , Boston, United States

Key Findings

- **Patients**

- N = 3
- Heavily pretreated RRMM (≥ 3 prior lines, high-risk cytogenetics)
- No prior BCMA-targeted therapy

- **Efficacy**

- 100% MRD-negative in bone marrow at month 1 (10^{-5} – 10^{-6} sensitivity)
- All achieved PR at month 1, deepening to VGPR by month 3
- Responses ongoing with no disease progression

- **CAR-T Expansion & Persistence**

- Robust *in vivo* T-cell expansion despite no lymphodepletion
- CAR⁺ cells up to 72% of CD3⁺ T cells at peak (~day 15)
- Memory-phenotype CAR-T cells detected in blood and BM through ≥ 3 months

- **Safety**

- Grade 2 CRS in 2/3 patients; no ICANS
- Limited cytopenias, no treatment-emergent infections
- Toxicity profile comparable to ex vivo CAR-T, with milder hematologic effects



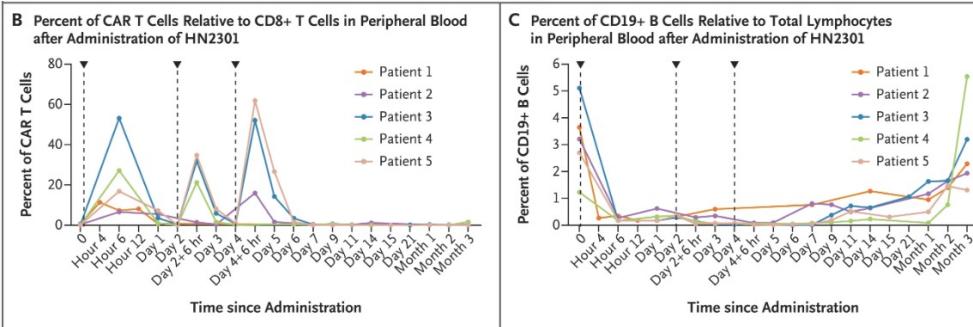
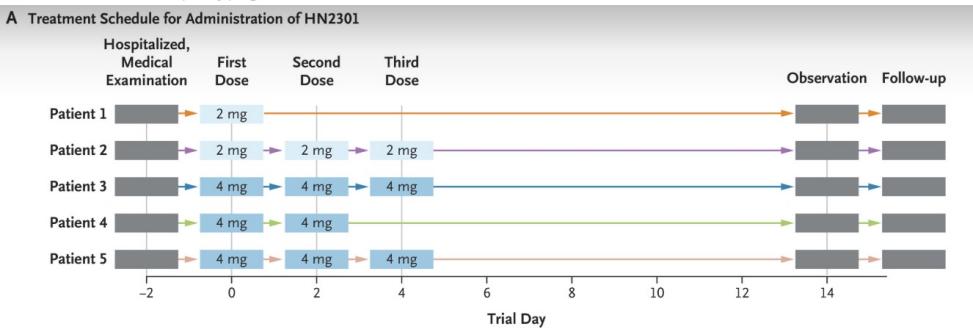
CORRESPONDENCE

f X in e

In Vivo CD19 CAR T-Cell Therapy for Refractory Systemic Lupus Erythematosus

Published September 17, 2025 | *N Engl J Med* 2025;393:1542-1544 | DOI: 10.1056/NEJMmc2509522

VOL. 393 NO. 15 | Copyright © 2025



Key Findings

- Rapid in vivo CAR-T generation:** CD19 CAR-T detectable within 6 h, transient expression (baseline by 2–3 days)
- Efficient B-cell depletion:** dose-dependent, complete depletion at 4 mg, lasting 7–10 days
- Favorable safety:** no Grade ≥ 3 CRS, no ICANS; only low-grade, manageable cytokine release
- Biologic & clinical activity:** reduced autoantibodies, normalized complement, SLEDAI-2K improved in all patients

Wang Q, et al. *N Engl J Med*. 2025;393(15):1542-1544.

Safety Challenges in In Vivo CAR-T Development

- **Limited clinical experience:** in vivo CAR-T remains early in development, requiring reliance on preclinical models and cautious clinical translation.
- **Platform-dependent risk profiles:** safety considerations differ fundamentally between delivery technologies
 - **Viral vector–based CARs:** genomic integration and persistent CAR expression limit control over expansion and durability, increasing risks of delayed inflammatory toxicities and chronic on-target effects.
 - **LNP–RNA–based CARs:** transient CAR expression may necessitate repeat dosing, introducing risks of innate immune activation, liver toxicity, and anti-vector immune responses.

Last Supper

